

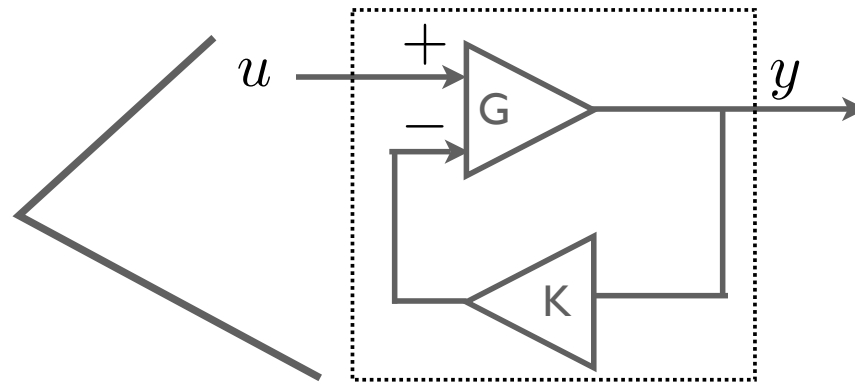
Design of in vitro Synthetic Gene Circuits



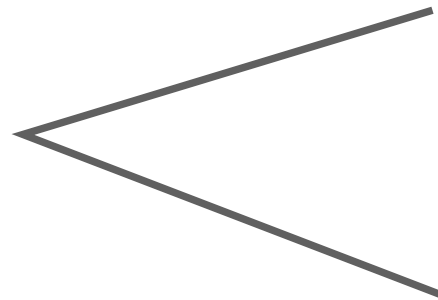
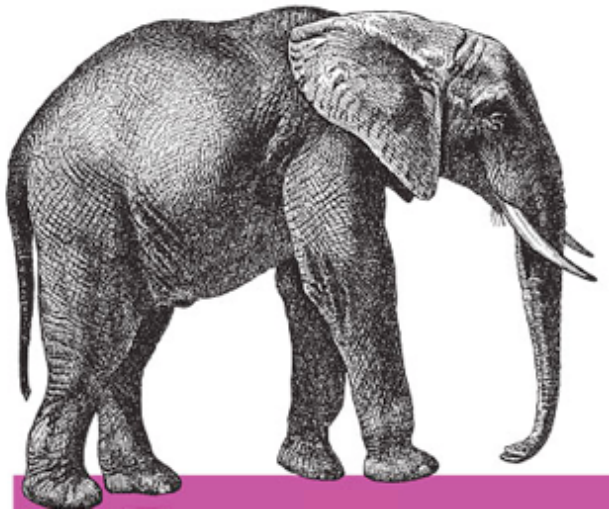
Elisa Franco
Richard M. Murray

15 June 2010
IWBDA

Programming large scale circuits



$$y = G(u - Ky)$$
$$y = u \frac{G}{1 + KG}$$
$$G \rightarrow \infty \Rightarrow y = \frac{1}{K}u$$



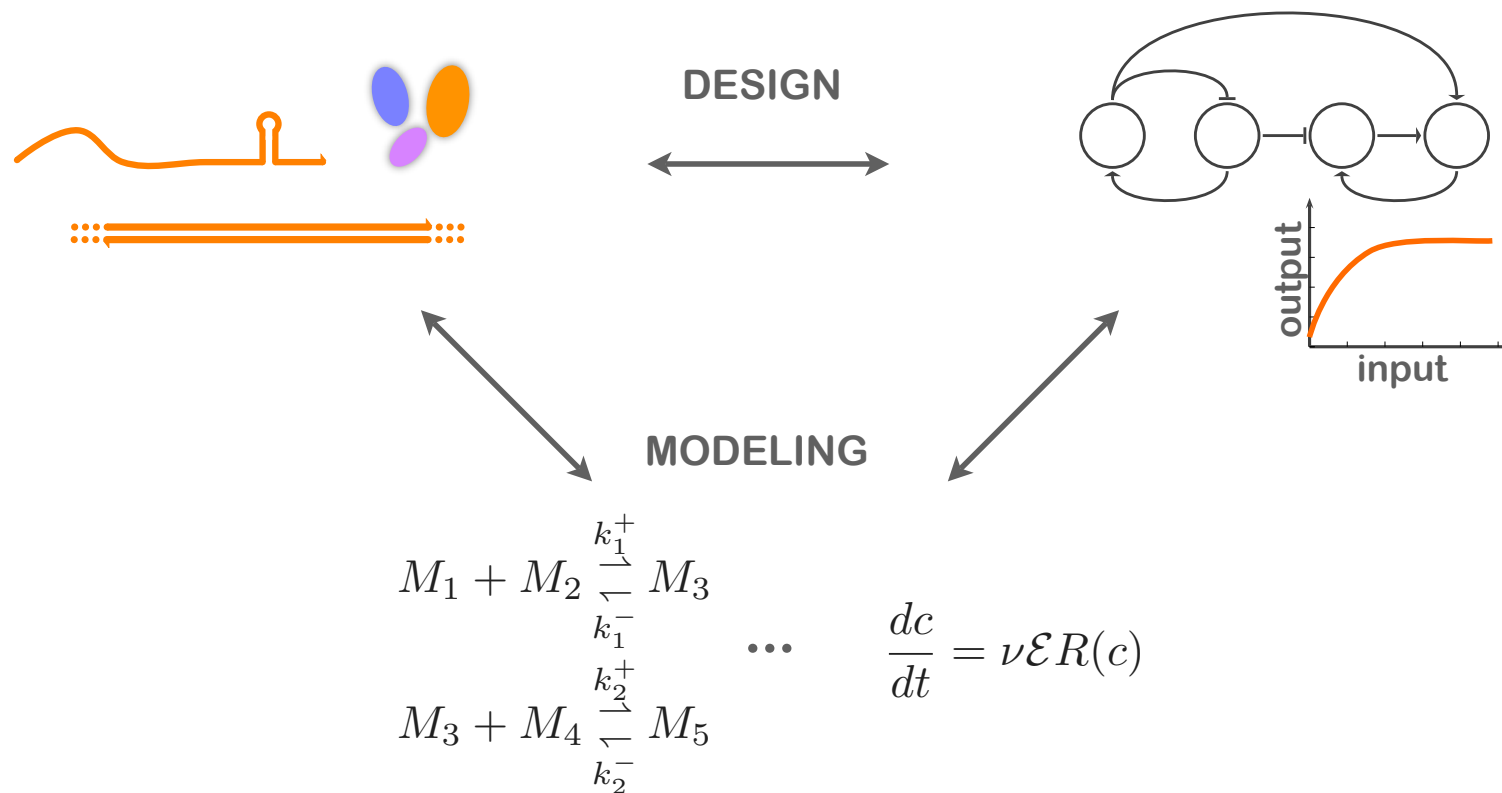
Bottom-up approach: In vitro molecular programming

OBJECTIVES:

Design and synthesis of biologically plausible feedback loops

Understand and implement design principles

Use few components: DNA and enzymes (off the shelf)



In this talk

In vitro genetic circuits: our tool kit

Programming a reaction network for rate regulation

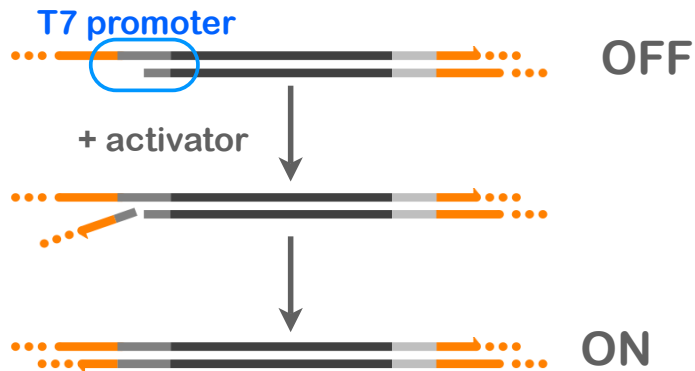
Interconnecting modules

Insulating devices

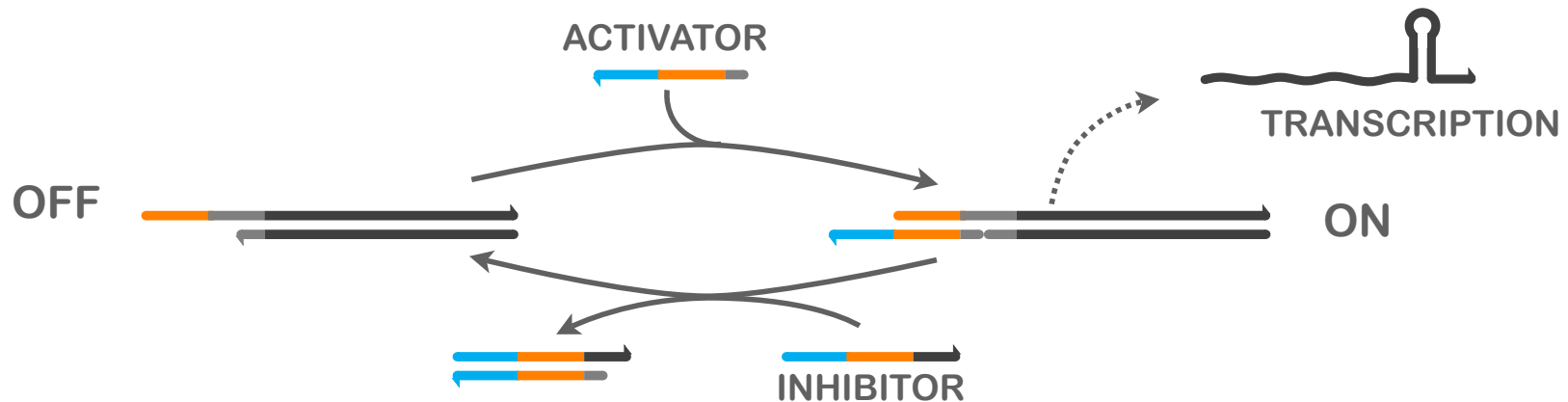
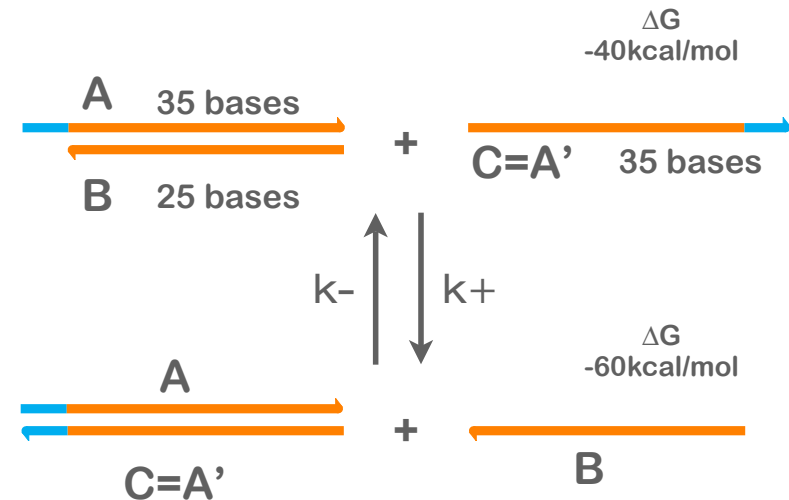
In vitro genetic circuits: the key ideas

Kim and Winfree Nature MSB06

Simplification of feedback loops:
Transcription is switched on and off
without transcription factors



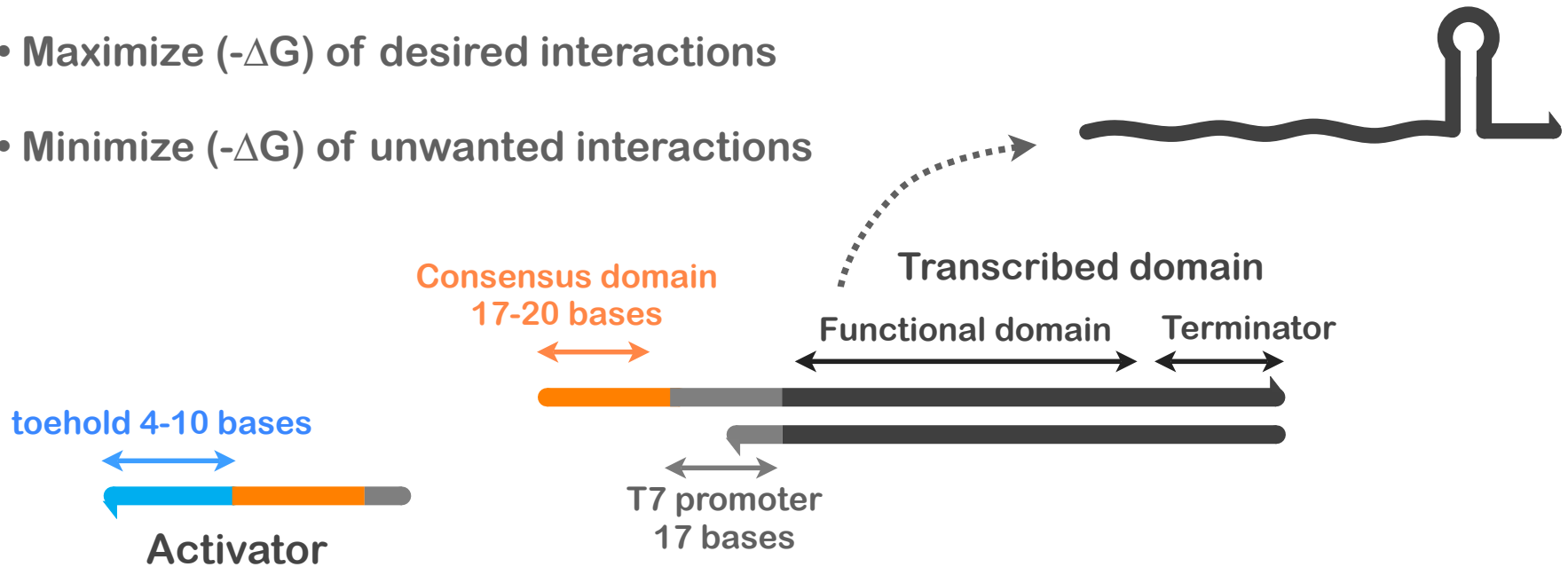
Switching: by toehold mediated branch migration - Yurke 03



Design and specification of parts

SOFTWARE:

- Maximize ($-\Delta G$) of desired interactions
- Minimize ($-\Delta G$) of unwanted interactions



Enzymes off the shelf:



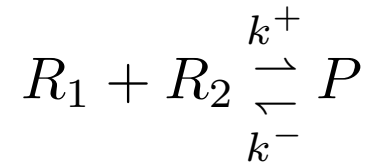
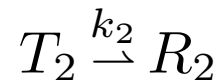
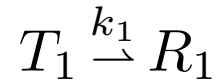
T7 RNA polymerase



RNase H

Programming a simple biochemical network

Produce reactants R1, R2
R1, R2 >> output P



Objective: steady flow of P

Constraints: avoid bottlenecks and waste of resources

IDEAS:

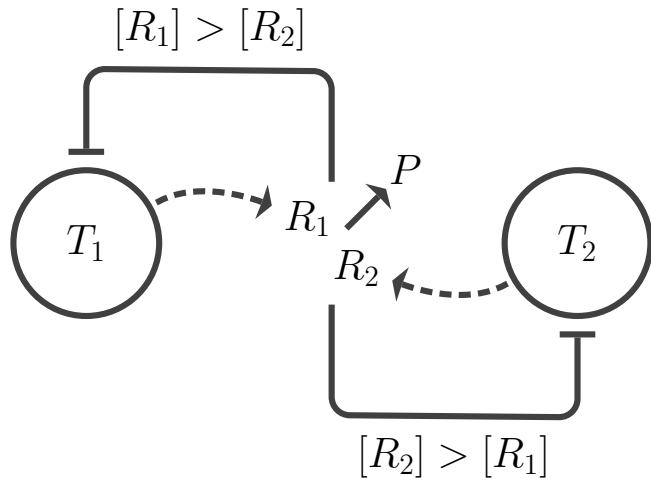
Negative feedback

Design network to decrease
excess species

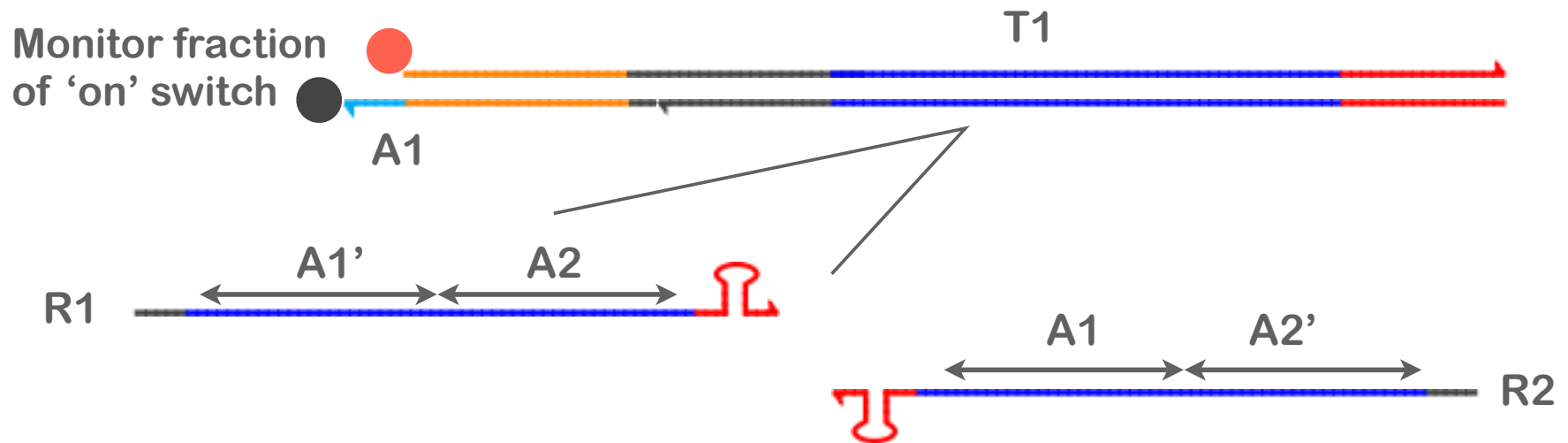
Positive feedback

Design network to stimulate
production of less abundant
species

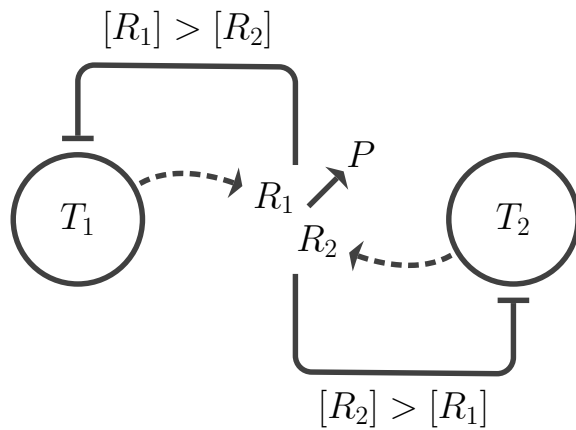
Self repression based flow regulation



- Transcriptional circuits implementation
- Reactants: transcripts
- Product: RNA complex
- **Circuit design:**



Modeling the dynamics



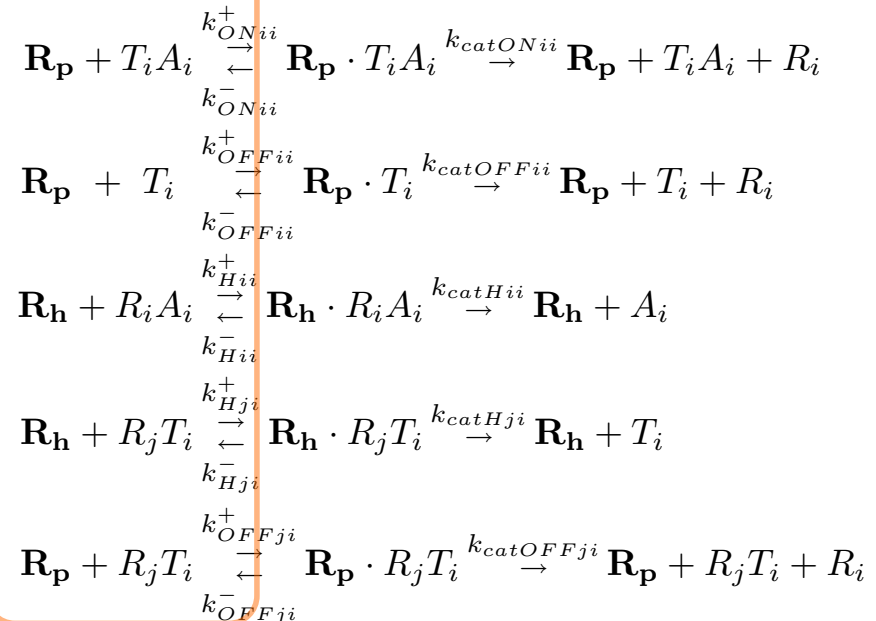
Mass action kinetics

Activation	$T_i + A_i \xrightarrow{k_{T_i A_i}} T_i A_i$
Inhibition	$R_i + A_i \xrightarrow{k_{R_i A_i}} R_i A_i$
	$R_i + T_i A_i \xrightarrow{k_{R_i T_i A_i}} R_i A_i + T_i$
Output	$R_i + R_j \xrightarrow{k_{R_i R_j}} R_i R_j$
Unwanted interactions	$R_j + T_i \xrightarrow{k_{R_j T_i}} R_j T_i$

Enzymatic reactions:

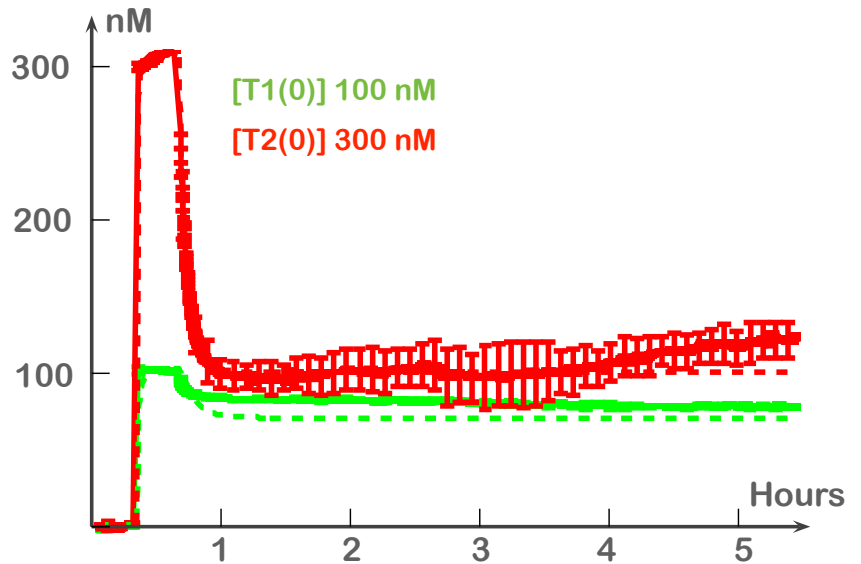
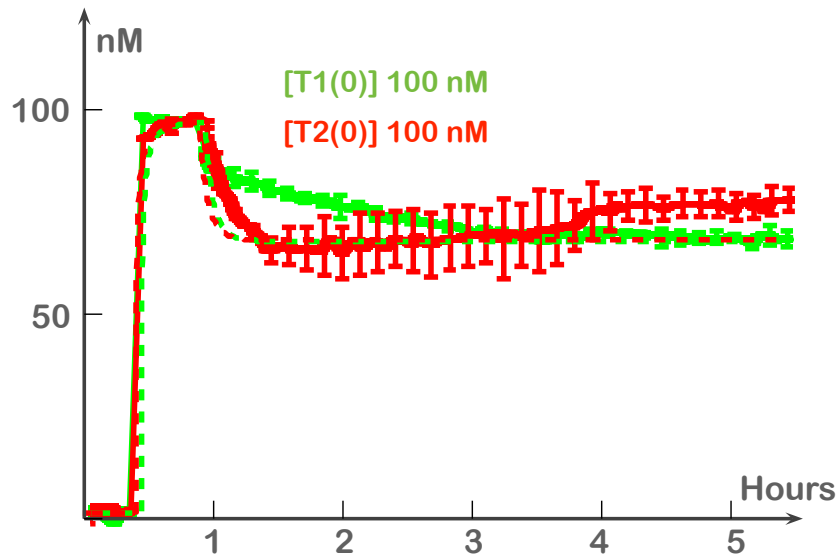
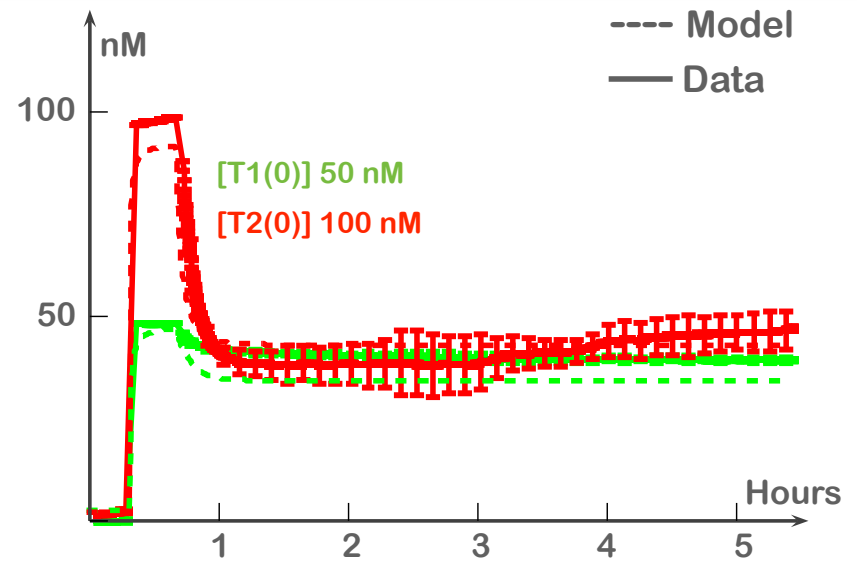
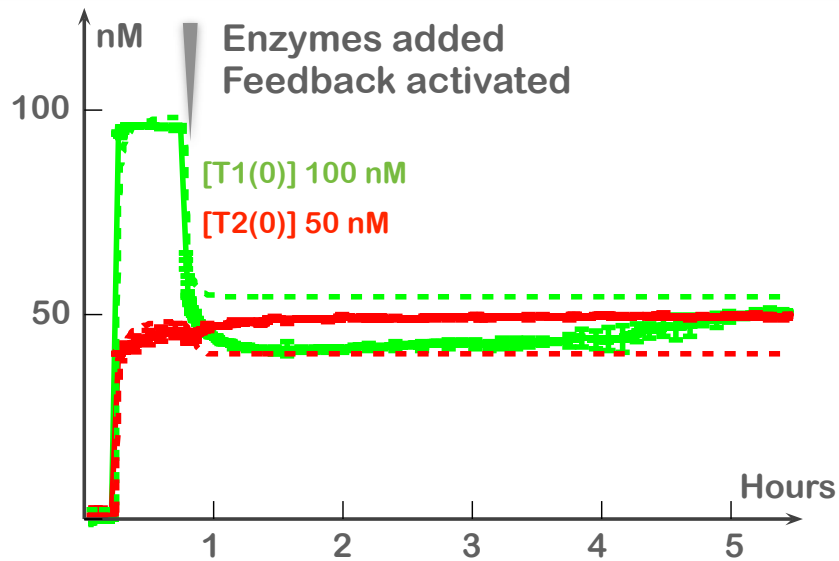
RNA polymerase - production of transcripts
RNase H - degradation of DNA/RNA hybrids

Michaelis-Menten kinetics

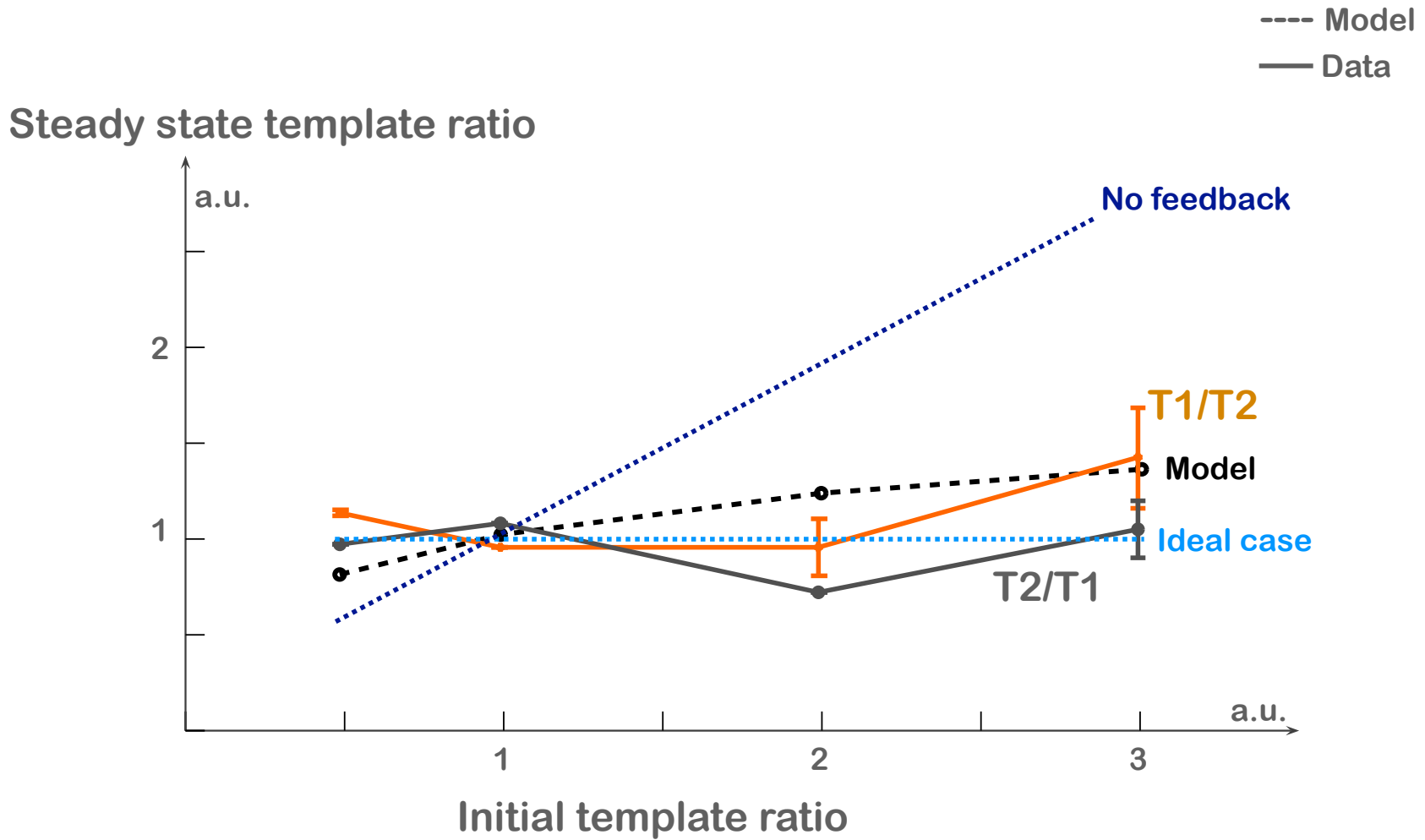


Faster

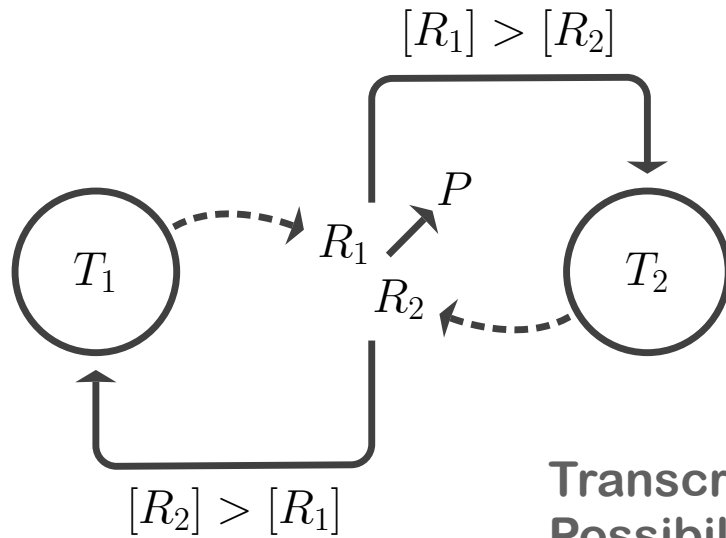
Experimental results



Ratio plot



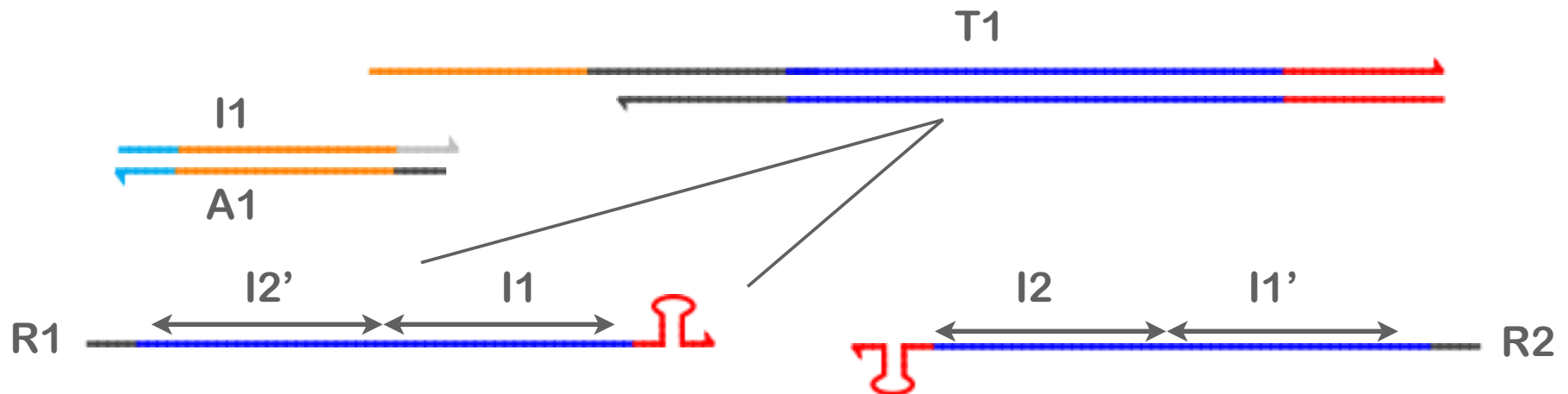
Cross activation circuit design



Reactants: transcripts

Product: RNA complex

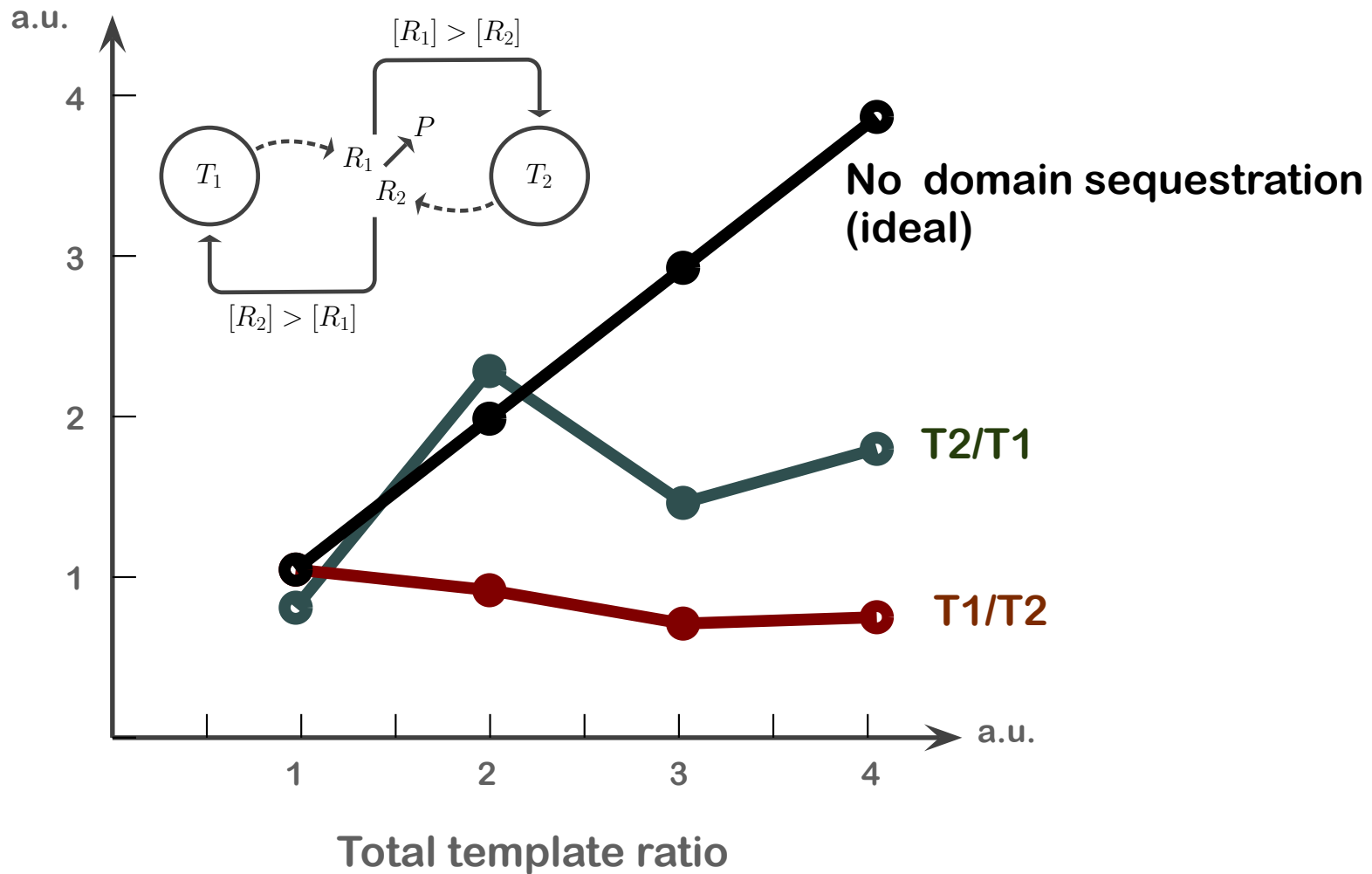
Transcripts: 'releaser' strands for the activators.
Possibility of unwanted self inhibition.



Preliminary data

Current design is asymmetric

Steady state 'on' template ratio



In this talk

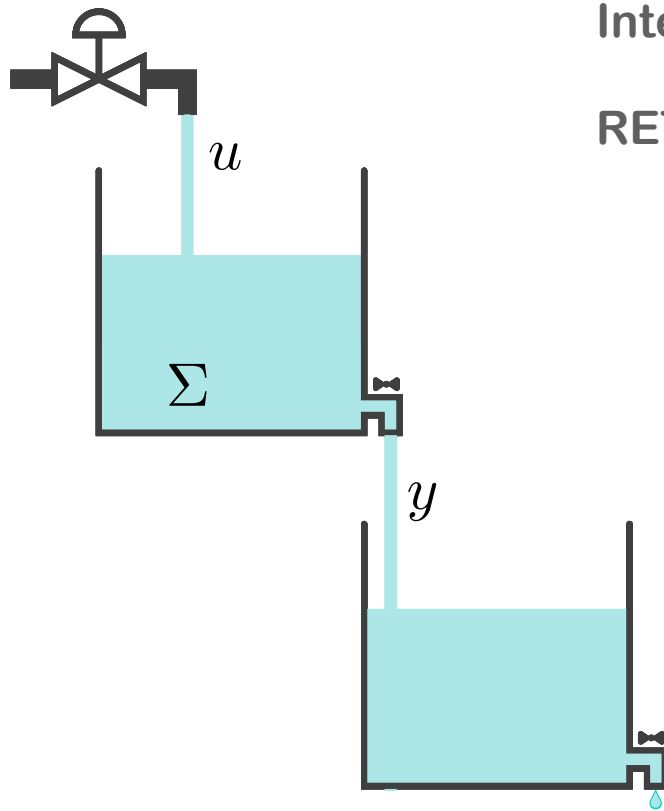
In vitro genetic circuits: our tool kit

Programming a reaction network for rate regulation

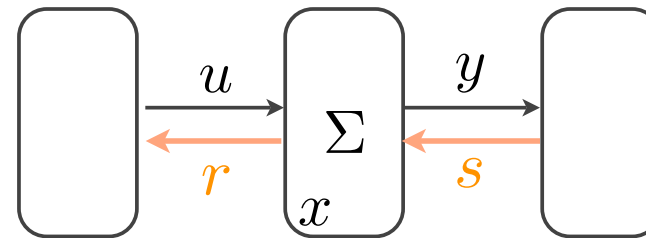
Interconnecting modules

Insulating devices

Interconnections can introduce unwanted dynamics



Interconnections introduce parasitic signals
RETROACTIVITY TO INPUTS AND OUTPUTS



$$\dot{x} = f(x, u, s)$$

$$y = Y(x, u, s)$$

$$r = R(x, u, s)$$

Del Vecchio et al. Nature MSB 2008

Existing theoretical results suggest how to design an insulating device

Del Vecchio, Ninfa and Sontag, MSB08; Del Vecchio, Jayanthi, ACC08

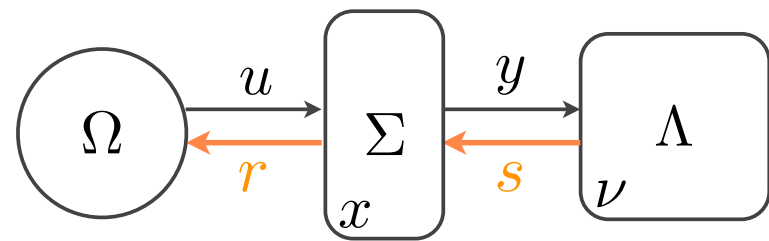
Structural assumptions

$$\Sigma : \begin{aligned} \dot{x} &= f(x, u, s) \\ y &= Y(x, u, s) \\ r &= R(x, u, s) \\ x &= (x_1, \dots, x_n) \in \mathcal{D} \subseteq \mathbb{R}_+^n \end{aligned}$$

1) u **positive scalars**
 $y = x_n$

2) $\Omega : \dot{u} = f_0(t, u)$ **prior to the interconnection**

3) $\Sigma : \dot{x} = \begin{pmatrix} Gf_1(x, u) \\ Gf_2(x) \\ \vdots \\ Gf_{n-1}(x) \\ Gf_n(x) \end{pmatrix}$



4) $\Lambda : \dot{\nu} = \begin{pmatrix} g_1(\nu, y) \\ g_2(\nu) \\ \vdots \\ g_p(\nu) \end{pmatrix}$

5) **Parasitic signals are additive**

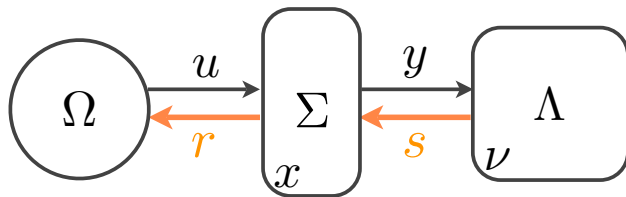
$$\begin{aligned} \dot{u} &= f_0(t, u) + r(x, u) \\ \dot{x}_n &= \dot{y} = Gf_n(x) + s(\nu, y) \end{aligned}$$

6) **Conservation laws**

$$\begin{aligned} r(x, u) &= -Gf_1(x, u) \\ s(\nu, y) &= -g_1(\nu, y) \end{aligned}$$

Existing theoretical results suggest how to design an insulating device

Stability assumption:



$$\Sigma : \begin{aligned} \dot{x} &= f(x, u, s) \\ y &= Y(x, u, s) \\ r &= R(x, u, s) \end{aligned}$$

$$\dot{x} = \begin{pmatrix} Gf_1(x, u) \\ Gf_2(x) \\ \vdots \\ Gf_{n-1}(x) \\ Gf_n(x) \end{pmatrix}$$

Define:

$$F : \mathbb{R}_+ \times \mathcal{D} \rightarrow \mathbb{R}^n$$

$$F(a, x) = (f_1(x, a - x_1), f_2(x), \dots, f_n(x))$$

$$a \in \mathbb{R}_+$$

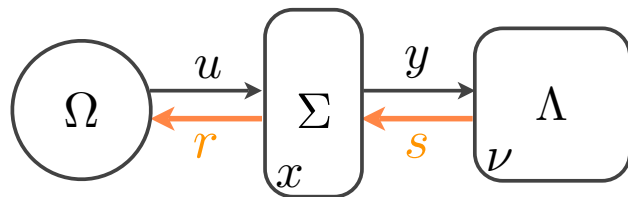
$$x \in \mathcal{D}$$

The Jacobian:

$$DF_x(a, x)$$

has all eigenvalues with negative real part in all its domain

The insulation property is achieved if the device is sufficiently fast



$$\Sigma : \begin{aligned} \dot{x} &= f(x, u, s) \\ y &= Y(x, u, s) \\ r &= R(x, u, s) \end{aligned}$$

$$\dot{x} = \begin{pmatrix} Gf_1(x, u) \\ Gf_2(x) \\ \vdots \\ Gf_{n-1}(x) \\ Gf_n(x) \end{pmatrix}$$

Claim 1

There exist G^* sufficiently large such that for any $G > G^*$:

$$\|x^{\text{ref}}(t) - x(t)\| = \mathcal{O}(1/G)$$

Where $x^{\text{ref}}(t)$ is the state of the device when the load is absent, i.e. $s(\nu, y) = 0$

Claim 2

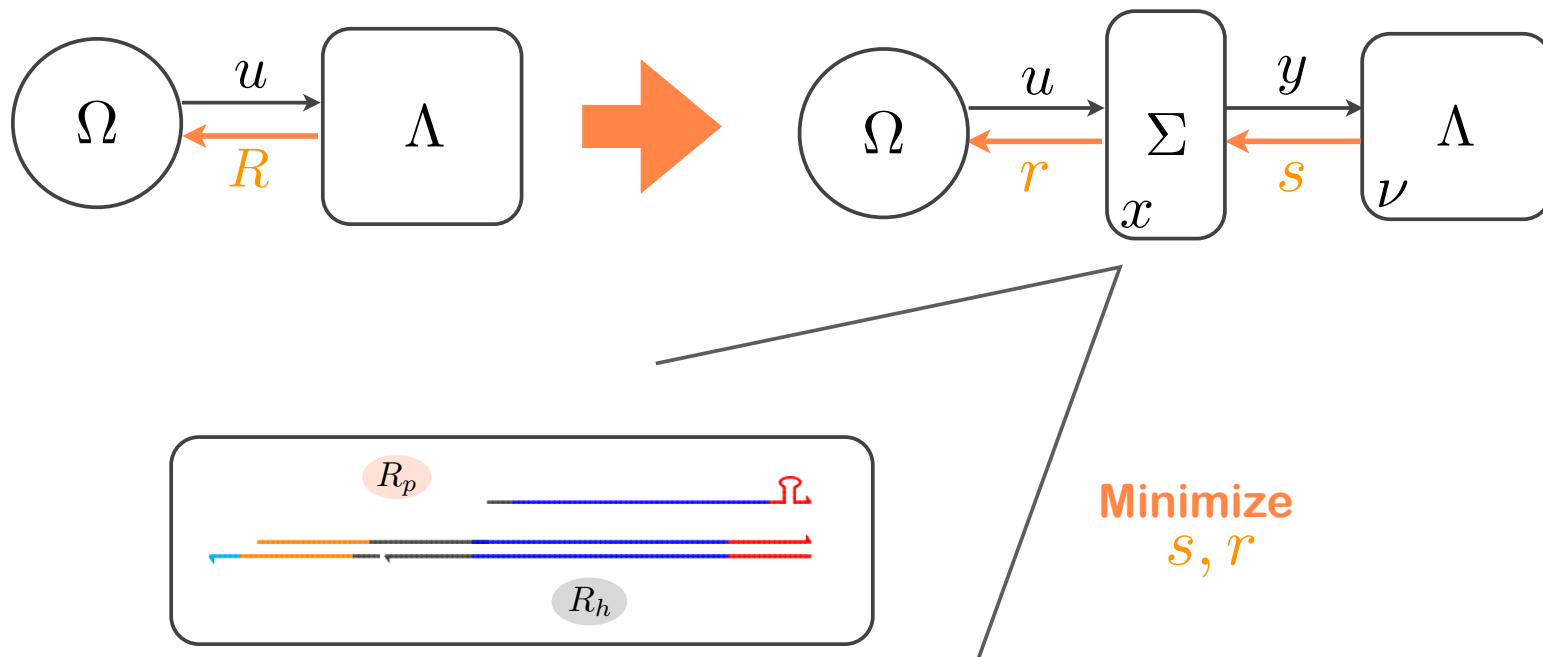
There exist G' sufficiently large such that for any $G > G'$:

$$u(t) = \bar{u}(t) + \mathcal{O}(1/G) \quad \text{where} \quad \frac{d\bar{u}}{dt} = f_0(t, u) \left(\frac{1}{(1 + \partial\gamma_1(\bar{u})/\partial\bar{u})} \right)$$

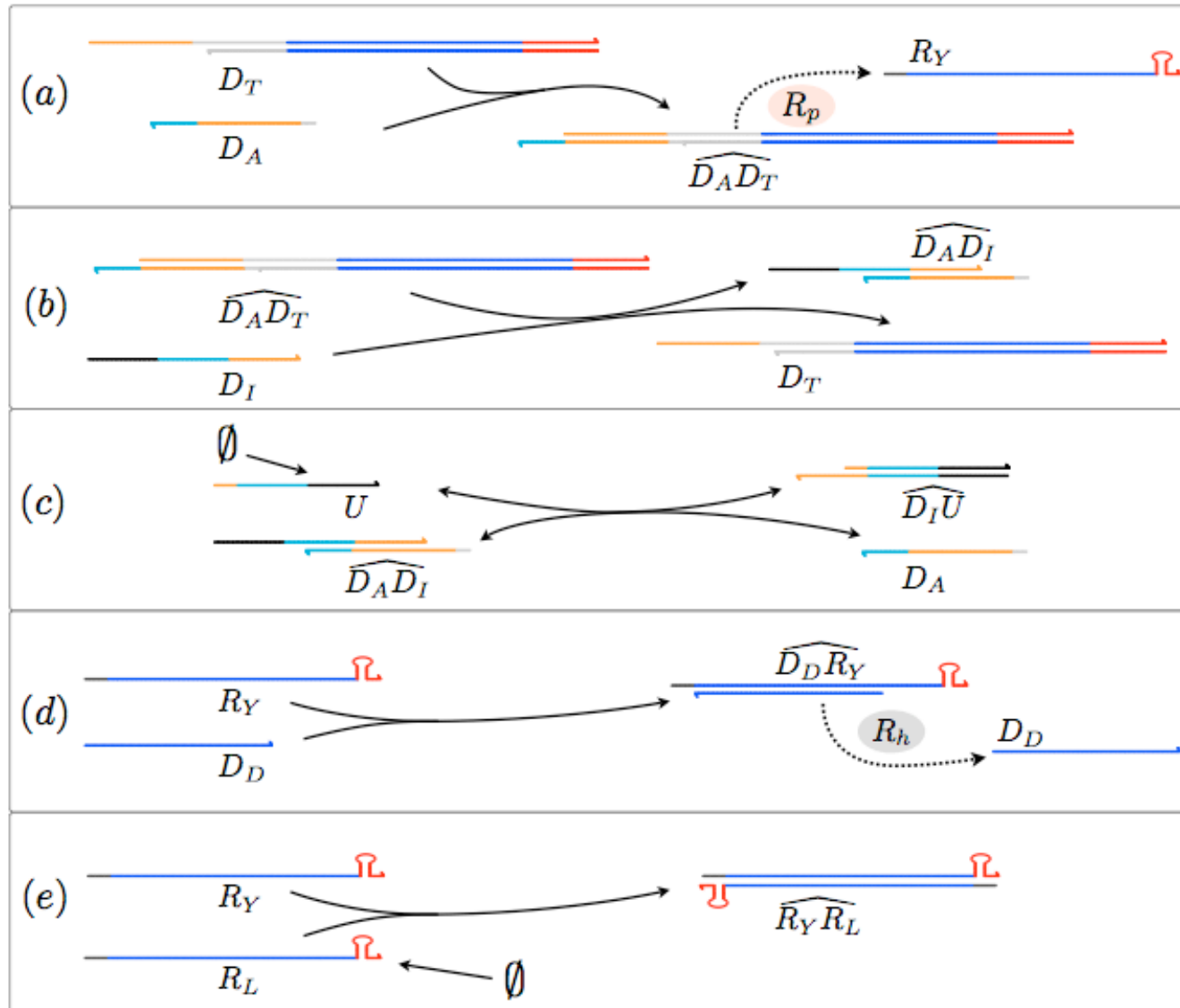
Low output retroactivity

All proofs are based on timescale separation

Can we make an insulator in an in vitro setting?

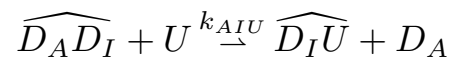
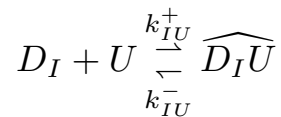
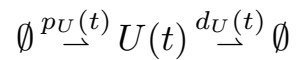


- Input U drives a switch
- RNA output binds to RNA load

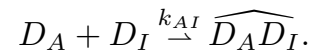
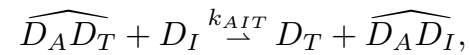
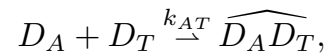


Reactions for a transcriptional insulator

Input stage



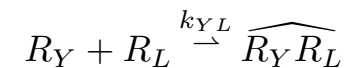
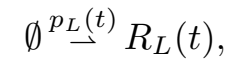
Core device



$$p_{R_Y}(t) = \alpha h \left(\frac{[\widehat{D_A D_T}]}{K_{MP}} \right)$$

$$d_{D_D R_Y}(t) = \gamma h \left(\frac{[\widehat{D_D R_Y}]}{K_{MH}} \right)$$

Output stage



Structural assumptions: dynamic gain can be tuned

Input

$$\dot{U} = +p_U(t) - d_U(t) - k_{IU}^+ D_I U + k_{IU}^- \widehat{D_I U} - k_{AIU} \widehat{D_A D_I U}$$

$$\widehat{D_I U} = +k_{IU}^+ D_I U - k_{IU}^- \widehat{D_I U} + k_{AIU} \widehat{D_A D_I U}$$

$$\widehat{D_A D_I} = +k_{AI} D_A D_I - k_{AIU} \widehat{D_A D_I U}$$

$$\widehat{D_A D_T} = +k_{AT} D_A D_T - k_{AIT} D_I \widehat{D_A D_T}$$

$$\dot{D} = -k_{DY} D_D R_Y + \gamma h \left(\frac{\widehat{D_D R_Y}}{K_{MH}} \right)$$

$$\dot{R}_Y = +\alpha h \left(\frac{\widehat{D_A D_T}}{K_{MP}} \right) - k_{DY} D_D R_Y - k_{YL} R_Y R_L$$

$$\dot{R}_L = +p_L(t) - k_{YL} R_Y R_L$$

Output

Core device

- Fast toehold kinetics
- High enzyme concentrations/activities

$$\dot{x} = \begin{pmatrix} G f_1(x, u) \\ G f_2(x) \\ \vdots \\ G f_{n-1}(x) \\ G f_n(x) \end{pmatrix}$$

Structural assumptions: additive retroactivity and conservation laws

Input

$$\dot{U} = +p_U(t) - d_U(t) - k_{IU}^+ D_I U + k_{IU}^- \widehat{D_I U} - k_{AIU} \widehat{D_A D_I U}$$

$$\widehat{D_I U} = +k_{IU}^+ D_I U - k_{IU}^- \widehat{D_I U} + k_{AIU} \widehat{D_A D_I U}$$

$$\widehat{D_A D_I} = +k_{AI} D_A D_I - k_{AIU} \widehat{D_A D_I U}$$

$$\widehat{D_A D_T} = +k_{AT} D_A D_T - k_{AIT} D_I \widehat{D_A D_T}$$

$$\dot{D}_D = -k_{DY} D_D R_Y + \gamma h \left(\frac{\widehat{D_D R_Y}}{K_{MH}} \right)$$

$$\dot{R}_Y = +\alpha h \left(\frac{\widehat{D_A D_T}}{K_{MP}} \right) - k_{DY} D_D R_Y - k_{YL} R_Y R_L$$

$$\dot{R}_L = +p_L(t) - k_{YL} R_Y R_L$$

Output

$$\dot{u} = f_0(t, u) + r(x, u)$$

$$\dot{x} = \begin{pmatrix} G f_1(x, u) \\ G f_2(x, u) \\ G f_3(x) \\ G f_4(x) \\ G f_5(x) + G s(\nu, y) \end{pmatrix}$$

$$\dot{\nu} = f_\nu(t) + G s(\nu, y).$$

Core device

Structural assumptions: stability

Jacobian of the dynamics of the core device:

$$DF_x(a, x) = \begin{bmatrix} P & \emptyset \\ L & Q \end{bmatrix} \quad \text{Lower diagonal}$$

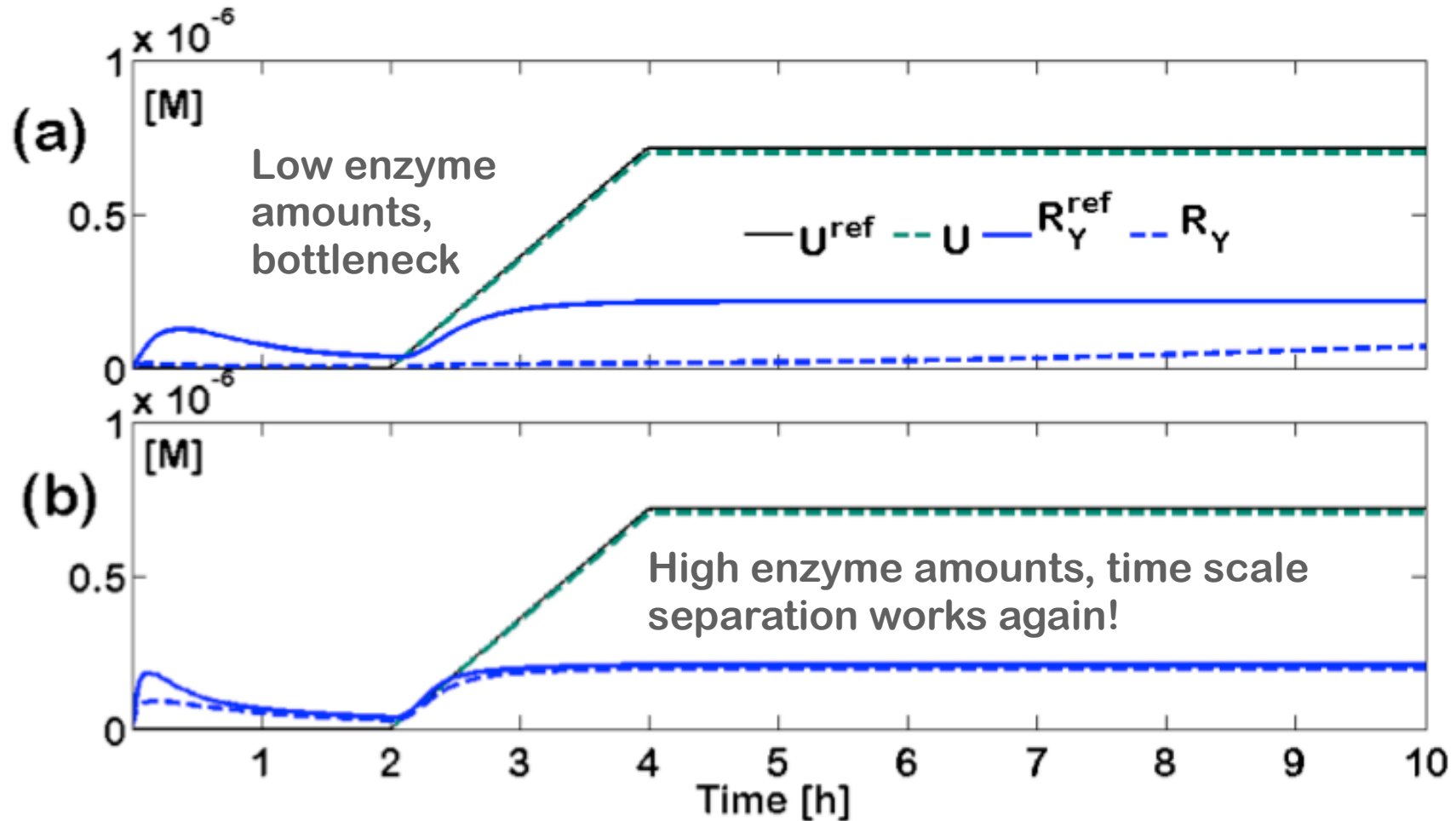
The eigenvalues have negative real part at any equilibrium point.

All structural assumptions are verified.

- Claim 1 holds: Low output retroactivity
- Claim 2 holds: Low input retroactivity

Note: Analytical mapping I/O not available, device may work in non linear regime

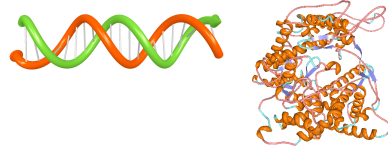
Simulation results



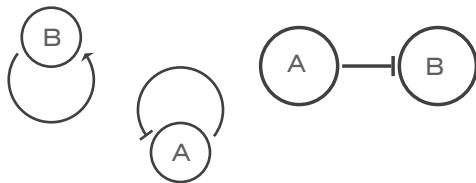
Programming synthetic biomolecular systems: embedding engineering principles in the hardware of life

Designing and building biosynthetic systems is today

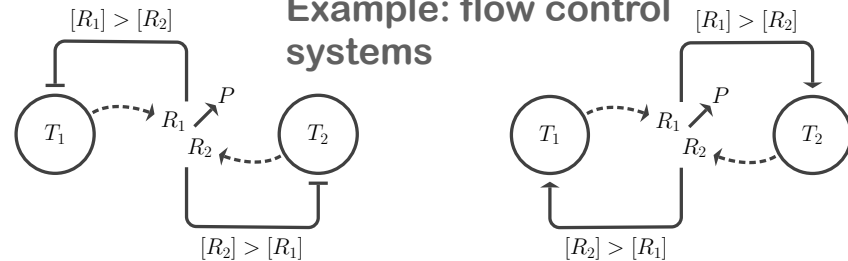
- Easier
- Faster
- Cheaper



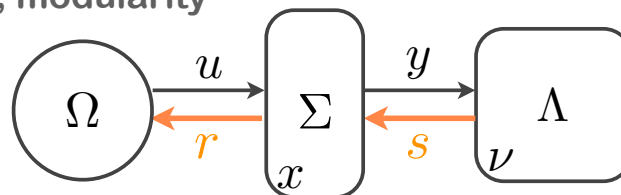
- In vitro bio-computational networks



Example: flow control systems



- Scaling up networks, modularity



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TU Munich

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